Lupin seeds lower plasma lipid concentrations and normalize antioxidant parameters in rats

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SUMMARY

Lupin seeds lower plasma lipid concentrations and normalize antioxidant parameters in rats. This study was designed to test bitter and sweet lupin seeds for lipid-lowering and for their antioxidative activities in hypercholesterolemic rats. The levels of plasma lipid, malondialdehyde (MDA) and whole blood reduced glutathione (GSH), as well as the activities of transaminases (ALT and AST), lactate dehydrogenase (LDH) in plasma, superoxide dismutase (SOD), glutathione peroxidase (GPx) in erythrocytes and plasma glutathione reductase (GR), glutathione-S-transferase (GST) and catalase (CAT) were examined. A hypercholesterolemia-induced diet manifested in the elevation of total lipids (TL), total cholesterol (TC), triglycerides (TG), LDL-C and MDA levels, ALT, AST, LDH activities and the depletion of GSH and enzymic antioxidants. The supplementation of a hypercholesterolemic-induced diet with bitter and sweet lupin seeds significantly lowered the plasma levels of TL, TC, TG and LDL-C. ALT, AST and LDH activities slightly decreased in treated groups compared with the hypercholesterolemic group (HC). Furthermore, the content of GSH significantly increased while MDA significantly decreased in treated groups compared with the HC group. In addition, the bitter lupin seed group improved enzymic antioxidants compared with the HC group. In general, the results indicated that the bitter lupin seed supplements are better than those containing sweet lupin seeds. These results suggested that the hypocholesterolemic effect of bitter and sweet lupin seed supplements might be due to their abilities to lower the plasma cholesterol level as well as to slow down the lipid peroxidation process and to enhance the antioxidant enzyme activity.


1. INTRODUCTION

Legume seeds are an abundant source of proteins and, among them, lupin is one of the richest. Indeed, lupin seed deserves greater interest as a result of its chemical composition and increased availability in many countries in recent years. Lupin is a non-starch leguminous seed with high protein content, almost as high as that of soybean (about 35% of the dry weight), relatively low oil content (Duranti et al., 2008) and a lack of antinutritional substances. The amount of antinutritional compounds found in other legumes, such as alkaloids, saponins, tannins and trypsin inhibitor, is minimal in lupin (Van Barneveld, 1999). Lupin cultivation is at least 2,000 years old and most likely began in Egypt or in the general Mediterranean region. The lupin plant, like other grain legumes (beans, peas, lentils, etc.) fixes atmospheric nitrogen, and produces seeds high in protein. There are over 300 species of the genus Lupinus (L.) (Putnam et al., 1989). Seeds of several species of lupins have been used as food for 3,000 years in the Mediterranean area (Gladstones, 1998). These bitter seeds had to be soaked in water...
before consumption, to remove most of their alkaloid content (Pettersson, 1998). From the second half of the 20th century onward, low-alkaloid varieties of white lupin (*Lupinus albus*), yellow lupin (*Lupinus luteus*), and blue lupin (*Lupinus angustifolius*) have been domesticated and selected (Cowling et al., 1998). In 2004, sweet varieties of these three species were mainly cultivated in several parts of Australia, Europe, and South America (Martins et al., 2005) and used for animal feed and food applications (Sirtori et al., 2010).

Hypercholesterolemia and its implications in cardiovascular diseases is a major problem in human health, and much attention has been paid to dietary intervention as a tool for its prevention and treatment (Kerckhoffs et al., 2002). Legumes have shown hypocholesterolemic effects in human and animal models (Martins et al., 2004), but these studies have mainly been done with soybean or its components. Therefore, studies involving other legumes, such as lupins, may clarify the mechanism by which plasma cholesterol is reduced and lead to the identification of new functional foods and/or components (Martins et al., 2005). There is limited information on the physiological effects of lupin seeds, particularly on the lipid metabolism. Viveros et al. (2007) reported a lower concentration of plasma cholesterol and triglycerides when lupins (*Lupinus albus*) were included in chicken diets. In addition, several studies have shown that this legume is characterized by hypocholesterolaemic (Betzzieche et al., 2008; Sirtori et al., 2004 and Spielmann et al., 2007), anti-atherogenic (Marchesi et al., 2008), hypotensive (Lee et al., 2008 and Pilvi et al., 2006), and hypoglycaemic activities (Hall et al., 2005a and Lee et al., 2006).

The present study was designed to compare the effect of bitter and sweet lupin seeds in protecting experimental animals fed a hypercholesterolemia-induced diet of oxidative stress and hypercholesterolemia.

2. MATERIALS AND METHODS

2.1. Materials

The seeds of bitter and sweet lupin varieties (*Lupinus albus*) were purchased from the National Research Center (NRC), Ministry of Agriculture, Giza, Egypt. The seeds were milled well into fine powders.

2.2. Animals and diets

Twenty-four male Sprague-Dawley rats weighing 100 ± 10 g were purchased from the animal house of Helwan Station for Experimental Animals, Helwan, Egypt. They were raised in the animal house of the Biochemistry Department, Faculty of Agriculture, Cairo University, Giza, Egypt. The animals were housed in polyethylene cages in groups of six rats per cage in a controlled environment (25±2°C, 50-60% relative humidity and 12-hour light-dark cycle). The animals were fed *ad libitum* with a basal diet and water for two weeks and were then randomly assigned to 4 groups (6 rats each): normal control group (NC), receiving basal diet consisting of corn starch 65%, casein 10%, corn oil 10%, salt mixture 4%, vitamin mixture 1% and cellulose 10% (AOAC,2000), high-cholesterol control group (HC) receiving hypercholesterolemia-induced diet which was prepared in the same way as the basal diet, except that the 10% corn oil portion was replaced with 10% sheep perineal fat and it was supplemented with 1% cholesterol and 0.25% cholic acid (Fukushima et al., 1997). The bitter lupin group was fed hypercholesterolemia-induced diets supplemented with 5% bitter lupin seeds. The sweet lupin group received hypercholesterolemia-induced diets supplemented with 5% sweet lupin seeds.

2.3. Experimental design

During the experimental period (4 weeks), water and diets were available *ad libitum*. At the end of the experiment, all the animals were sacrificed by decapitation. Blood samples were collected in three heparinized tubes. The first one (0.1 ml blood) was used for the determination of reduced glutathione (GSH), the second heparinized tube (0.5 ml blood) was used to extract the erythrocytes lysate according to the procedure of Quist (1980) in order to study antioxidant enzymes. The third heparinized tube was centrifuged at 2500 rpm at 37°C for 15 min to separate the plasma.

2.4. Biochemical analysis

**Lipid analysis**

The total lipids in plasma (TL), triglycerides (TG), total cholesterol (TC), low density lipoprotein cholesterol (LDL-C) and high density lipoprotein cholesterol (HDL-C) were determined according to Allain et al. (1974), Levy (1981) and Burstein (1970), respectively. The atherogenic Index (AI) was calculated according to Lee and Niemann (1996) using following equation:

\[
\text{Atherogenic Index (AI)} = \frac{\text{Total cholesterol} - \text{HDL-C}}{\text{HDL-C}}
\]

**Determination of LDH, AST and ALT activities**

The lactate dehydrogenase (LDH) activity in plasma was determined according to the method of Young (2001). Aspartate aminotransferase (AST) and alanine aminotransferase (ALT) activities were measured colorimetrically in plasma according to the method described by Reitman and Frankel (1957).
**Determination of glucose**

The plasma glucose level was determined according to Trinder (1969).

**Determination of lipid peroxidation**

Plasma lipid peroxidation was estimated by measuring the thiobarbituric acid reactive substances (TBARS) and was expressed in terms of malondialdehyde (MDA) content, according to the method of Uchiyama and Mihara (1978). The MDA values were estimated using 1,1,3,3-tetraethoxy propane as the standard.

**Determination of non-enzymic antioxidants (GSH)**

Reduced glutathione (GSH) in whole blood was determined by the method of Beutler et al. (1963). This method was based on the reaction of GSH with 5,5’-dithiobis(2-nitrobenzoic acid) to give a yellow compound that absorbs at 412 nm.

**Determination of enzymic antioxidant activities**

Superoxide dismutase (SOD) and glutathione peroxidase (GPx) in erythrocytes were assayed by the methods of Nishikimi et al. (1972) and Paglia and Valentine (1970), respectively. Plasma glutathione reductase (GR) activity was assayed by the method of Goldberg and Spooner (1983). The enzyme activity was quantified by measuring the disappearance of NADPH at 340 nm. Plasma glutathione-S-transferase (GST) activity was determined using the procedure of Habig et al. (1974) by measuring the conjugation of 1-chloro-2,4-dinitrobenzene (CDNB) with reduced glutathione. The conjugation is accompanied by an increase in absorbance at 340 nm. The rate of increase is directly proportional to the GST activity. Plasma catalase (CAT) activity was determined according to the method of Aebi (1984).

**Statistical analysis**

A statistical analysis (standard deviation “SD” and standard error “SE”) was carried out according to Fisher (1970). The LSD (Least significant difference) test was used to compare the significant differences between means of treatment (Waller and Duncan, 1969). The statistical package for social science S.P.S.S. (1999) program version was used for all analyses.

### Table 1

<table>
<thead>
<tr>
<th>Groups</th>
<th>Total lipids (mg/dl)</th>
<th>Total cholesterol (mg/dl)</th>
<th>Triglycerides (mg/dl)</th>
</tr>
</thead>
<tbody>
<tr>
<td>NC</td>
<td>285.02 ± 7.32</td>
<td>91.30 ± 2.33</td>
<td>93.66 ± 4.05</td>
</tr>
<tr>
<td>HC</td>
<td>1163.12 ± 48.30</td>
<td>487.70 ± 33.82</td>
<td>281.80 ± 16.45</td>
</tr>
<tr>
<td>Bitter lupin</td>
<td>601.49 ± 34.82</td>
<td>287.50 ± 33.17</td>
<td>204.33 ± 24.97</td>
</tr>
<tr>
<td>Sweet lupin</td>
<td>875.00 ± 36.23</td>
<td>321.59 ± 26.34</td>
<td>268.59 ± 29.69</td>
</tr>
<tr>
<td>LSD</td>
<td>107.98</td>
<td>83.58</td>
<td>65.23</td>
</tr>
</tbody>
</table>

Values are expressed as means ± SE (n = 6). Values with different superscript letters within the same column are significantly different (P < 0.05).

### RESULTS AND DISCUSSIONS

In the present study, hypercholesterolemia-induced diet feeding for four weeks was chosen as the experimental model of early phase atherogenesis. Cholic acid addition enhances the hypercholesterolemic effect of cholesterol feeding (Shinnick et al., 1990). The role of bitter and sweet lupin seeds in countering the lipidemic-oxidative aberrations accompanying diet-induced hypercholesterolemia have been investigated here.

Rats fed the hypercholesterolemia-induced diet (HC) developed hypercholesterolemia marked by a significant (P < 0.05) increase in plasma total lipids (TL), triglycerides (TG), total cholesterol (TC), low density lipoprotein cholesterol (LDL-C), high density lipoprotein cholesterol (HDL-C) and atherogenic Index (AI) compared with the normal control rats (NC). Supplementation with bitter and sweet lupin seeds showed significant (P < 0.05) falls in total lipids, triglycerides, total cholesterol, LDL-C and AI and a insignificant increase in HDL-C compared with the hypercholesterolemic group (HC) as shown in Tables (1 and 2). The best reduction in the lipids profile was recorded for the bitter lupin seed supplement; the levels of total lipids, total cholesterol, triglycerides, LDL-C and AI were decreased by 48.29%, 41.05%, 27.49%, 46.61%, and 54.22%, respectively. A significant increase in the HDL-C level was observed for the bitter lupin seed supplement. However, no significant change in the HDL-C level was observed for the sweet lupin seed supplement. The results show that bitter lupin seed supplemented diets are more effective against hypercholesterolemia than sweet lupin seeds. These results are in agreement with the observations of Rahman et al. (1996), Chango et al. (1998), Sirtori et al. (2004), Spielmann et al. (2007) and Bettziche et al. (2008) using lipid seeds in rats; Eder et al. (1996) and Rubio et al. (2003) in...
content supernatant viscosity is highly correlated with reduced serum and liver cholesterol (Gallaher et al., 1993) and reductions in cholesterol absorption (Carr et al., 1996) in hamsters. Increased bile acid excretion represents another mechanism by which a reduction in cholesterol can be produced. Costa et al. (1994) demonstrated that viscous NSP can enhance bile secretion and subsequently result in a significant loss of these acids in the feces of rats. The continued drain of bile acids and lipids by sequestration and increased elimination as fecal acidic and neutral esters may ultimately influence the absorption of lipids and cholesterol in the intestine. Although soluble dietary fiber was known to be an effective hypocholesterolemic agent, the insoluble dietary fiber of legume seeds has also been reported to be effective in lowering serum cholesterol in hypercholesterolemic men (Hughes, 1991). The addition of lupin kernel fiber to the diet provided favorable changes to some serum lipid (total cholesterol, high-density lipoprotein cholesterol) (Hall et al., 2005b). These studies confirm the results of the present study. In fact, bitter lupin seeds may prevent an increase in the factors causing coronary heart diseases (CHD) and cardiovascular diseases (CVD) more than sweet lupin seeds, so they may also prevent atherosclerosis.

Table (3) presents the results of plasma AST, ALT and LDH activities in the experimental rats. There were significant increases (P<0.05) in the plasma AST, ALT and LDH activities of hypercholesterolemic rats (HC) as compared to normal control rats (NC). The present findings are in agreement with those obtained by Ahmed et al. (1987) who found that hypercholesterolemia states significantly stimulate ALT and AST activity in the

<table>
<thead>
<tr>
<th>Groups</th>
<th>HDL-cholesterol</th>
<th>LDL-cholesterol</th>
<th>Al</th>
</tr>
</thead>
<tbody>
<tr>
<td>NC</td>
<td>38.33 ± 1.28abc</td>
<td>37.05 ± 3.30abc</td>
<td>1.38</td>
</tr>
<tr>
<td>HC</td>
<td>45.49 ± 2.62abc</td>
<td>370.99 ± 33.63abc</td>
<td>9.72</td>
</tr>
<tr>
<td>Bitter lupin</td>
<td>52.74 ± 2.93a</td>
<td>198.07 ± 29.73abc</td>
<td>4.45</td>
</tr>
<tr>
<td>Sweet lupin</td>
<td>37.58 ± 2.53c</td>
<td>235.27 ± 21.73abc</td>
<td>7.56</td>
</tr>
<tr>
<td>LSD</td>
<td>7.47</td>
<td>76.90</td>
<td></td>
</tr>
</tbody>
</table>

Values are expressed as means ± SE (n = 6). Values with different superscript letters within the same column are significantly different (P < 0.05).

<table>
<thead>
<tr>
<th>Groups</th>
<th>ALT</th>
<th>AST</th>
<th>LDH</th>
</tr>
</thead>
<tbody>
<tr>
<td>NC</td>
<td>22.35 ± 0.07d</td>
<td>25.22 ± 0.46c</td>
<td>560.5 ± 15.4c</td>
</tr>
<tr>
<td>HC</td>
<td>36.37 ± 0.53a</td>
<td>98.99 ± 6.63a</td>
<td>1232.89 ± 20.5a</td>
</tr>
<tr>
<td>Bitter lupin</td>
<td>28.14 ± 0.38c</td>
<td>61.17 ± 4.51b</td>
<td>963.7 ± 21.7b</td>
</tr>
<tr>
<td>Sweet lupin</td>
<td>31.16 ± 0.40b</td>
<td>70.35 ± 5.16b</td>
<td>1127.0 ± 74.5a</td>
</tr>
<tr>
<td>LSD</td>
<td>1.19</td>
<td>14.71</td>
<td>151.05</td>
</tr>
</tbody>
</table>

Values are expressed as means ± SE (n = 6). Values with different superscript letters within the same column are significantly different (P < 0.05).
plasma. Plasma ALT activity significantly decreased (P<0.05) in rats fed the hypercholesterolemia-induced diets supplemented with bitter or sweet lupin seeds by 22.63% and 14.32%, respectively compared with the hypercholesterolemic control (HC). Moreover, plasma AST activity significantly decreased in all treated groups as compared to the hypercholesterolemic control (HC). The highest decrease in plasma AST activity was recorded for the bitter lupin seed supplement (38.20%). No changes in LDH activity was observed for the sweet lupin seed group when compared with the hypercholesterolemic control group (HC). However, there was a significant decrease in the plasma LDH activity of the bitter lupin seed group as compared to the hypercholesterolemic group (HC).

Mansour et al. (2002) found that a treatment of alloxan-diabetic rats with *Lupinus albus* for 28 consecutive days could restore the activities of AST, ALT and LDH to their normal levels. A possible explanation for the effect of lupin seeds on the activities of AST, ALT and LDH in plasma and liver is that these seeds may inhibit the liver damage induced by hypercholesterolemia.

The liver is a central organ for many physiological and biochemical processes necessary for the maintenance of life (Souba and Wilmore, 1983). Morphological alterations that occur in the liver affect many metabolic processes in the organism. Peroxide formation induced by hypercholesterolemia (Sudhahar et al., 2007) results in the release of some enzymes by interacting with cellular structure and function. Thus, the serum activities of cellular enzymes such as transaminases, alkaline phosphatase, and lactate dehydrogenase do increase. With the increase in cellular membrane permeability, intracellular fluid transfers into intercellular space, resulting in muscle and liver cell degeneration.

In the present study, it was observed that as a result of hypercholesterolemia, enzymes such as AST, ALT and LDH were released into the blood. Their increase in the plasma activities of these enzymes was directly proportional to the degree of cellular damage. These values decreased with bitter and sweet lupin seed supplements.

Rats fed hypercholesterolemia-induced diets showed a significant (P<0.05) increase and a decrease in plasma malondialdehyde (MDA) level and blood reduced glutathione (GSH) content respectively compared to the normal control rats (Table 4). MDA significantly increased (P<0.05) by 203.63% and GSH significantly depleted (P<0.05) by 31.13% in hypercholesterolemic rats (HC). These results are related to the results of Kempaiah and Srinivasan (2004 and 2005). Supplementation of hypercholesterolemia-induced diets with bitter lupin seeds showed a significant (P<0.05) decrease and an increase in MDA level and GSH content respectively compared with hypercholesterolemic rats (Table 4). The best results of MDA and GSH were recorded for the bitter lupin seed supplement. MDA significantly decreased (P<0.05) by 40.16% and GSH significantly increased (P<0.05) by 44.50% in the bitter lupin seed group compared with hypercholesterolemic rats (HC). However, no significant changes in MDA and GSH levels were observed for the sweet lupin seed supplement.

MDA level is the most important factor for indicating an increased peroxidative level, while glutathione is a substance with an important role in cell detoxification and protection from hazardous compounds. Glutathione is synthesized in the erythrocytes and is found in living cells. It has been reported that cellular glutathione has an important function against chemical agents by protecting the cell membrane integrity. A decrease in the amount of glutathione and an increase in the amount of MDA may result in the destruction of membrane integrity (Kempaiah and Srinivasan, 2005 and Tauseef et al., 2007). In this study, the decrease in the reduced glutathione and the increase in malondialdehyde levels of hypercholesterolemic group indicate that hypercholesterolemia damaged the integrity of the erythrocyte membrane. On the contrary, the observed increase in the amount of GSH and the decrease in MDA in the bitter lupin seed group indicate that bitter lupin seeds effectively protect membrane integrity.

Bitter lupin seeds have a hypolipidemic effect which may prevent the increase in lipid peroxidatin, therefore depleting GSH in hypercholesterolemia. The study reported by Sheweita et al. (2002) showed that the level of the free radicals, TBARS, was decreased in the Termis-treated diabetic rats for 28 consecutive days. The maintenance of free radical levels in Termis-treated diabetic animals

### Table 4

<table>
<thead>
<tr>
<th>Groups</th>
<th>GSH (mg/dl)</th>
<th>MDA (nmol/L)</th>
<th>Glucose (mg/dl)</th>
</tr>
</thead>
<tbody>
<tr>
<td>NC</td>
<td>45.68 ± 3.44</td>
<td>29.98 ± 1.11</td>
<td>86.10 ± 2.93</td>
</tr>
<tr>
<td>HC</td>
<td>31.46 ± 1.07</td>
<td>91.03 ± 2.60</td>
<td>114.09 ± 2.77</td>
</tr>
<tr>
<td>Bitter lupin seeds</td>
<td>45.46 ± 3.29</td>
<td>54.47 ± 4.69</td>
<td>94.47 ± 3.21</td>
</tr>
<tr>
<td>Sweet lupin seeds</td>
<td>33.60 ± 3.60</td>
<td>93.46 ± 4.54</td>
<td>105.10 ± 5.69</td>
</tr>
<tr>
<td>LSD</td>
<td>9.83</td>
<td>10.96</td>
<td>12.56</td>
</tr>
</tbody>
</table>

Values are expressed as means ± SE (n = 6). Values with different superscript letters within the same column are significantly different (P < 0.05).
The plasma glucose level of hypercholesterolemic rats (HC) was significantly (P < 0.05) increased compared to the normal control (NC). However, a significant improvement in the plasma glucose level was observed for the bitter lupin seed treated group compared with the hypercholesterolemic group (HC). However, no significant change was observed for the sweet lupin seed group.

* * *  

Lupinus albus (Termis) exerted hypoglycemic effects and an increase in the level of serum insulin in normal and diabetic subjects as well as in normal and alloxan-diabetic animals (Youness et al., 1985; Eskander and Won Jun, 1995 and Mansour et al., 2002).

The reduction of serum glucose level in the current experiment, by a lupin supplemented diet, is in agreement with the in vitro and in vivo study between a lupin seed protein (namely, conglutin γ) and insulin, reported recently by Magni et al. (2004). The effect of the oral administration of conglutin γ on the glycemic levels of rats subjected to glucose overloading resulted in a significant reduction in rat glycemia.

Table (5) displays the activities of antioxidant enzymes in erythrocytes and plasma. The erythrocytes superoxide dismutase (SOD), plasma glutathione reductase (GR) and glutathione-S-transferase (GST) were significantly (P < 0.05) inhibited (24.51%, 24.49% and 24.25%, respectively) in hypercholesterolemic rats (HC). A slight inhibition in the activities of erythrocytes glutathione peroxidase (GPx) and plasma catalase (CAT) was observed in the hypercholesterolemic group (HC) compared to the normal control group (NC). Hypercholesterolemia-induced diets supplemented with bitter lupin seeds improved the activities of these enzymes. However, no significant changes were observed for sweet lupin seed supplements.

Table 5  

<table>
<thead>
<tr>
<th>Groups</th>
<th>SOD (U/ml)</th>
<th>GPx (U/L)</th>
<th>CAT (U/L)</th>
<th>GR (U/L)</th>
<th>GST (U/L)</th>
</tr>
</thead>
<tbody>
<tr>
<td>NC</td>
<td>180.76 ± 7.17a</td>
<td>346.36 ± 31.36a</td>
<td>422.73 ± 30.83a</td>
<td>36.67 ± 3.93a</td>
<td>41.82 ± 2.08a</td>
</tr>
<tr>
<td>HC</td>
<td>136.46 ± 3.13a</td>
<td>310.27 ± 17.89a</td>
<td>377.74 ± 31.36a</td>
<td>27.69 ± 1.73bc</td>
<td>31.68 ± 2.73b</td>
</tr>
<tr>
<td>Bitter lupin</td>
<td>164.68 ± 8.54ab</td>
<td>341.26 ± 32.43a</td>
<td>384.18 ± 27.23a</td>
<td>34.16 ± 0.96bc</td>
<td>37.65 ± 2.18ab</td>
</tr>
<tr>
<td>Sweet lupin</td>
<td>148.43 ± 5.02abc</td>
<td>313.93 ± 14.09ab</td>
<td>353.11 ± 20.11b</td>
<td>24.62 ± 1.97bc</td>
<td>31.48 ± 1.69b</td>
</tr>
<tr>
<td>LSD</td>
<td>24.71 ± 6.32</td>
<td>67.32 ± 9.03</td>
<td>90.37 ± 6.91</td>
<td>6.91 ± 1.69</td>
<td>8.61 ± 1.69</td>
</tr>
</tbody>
</table>

Values are expressed as means ± SE (n = 6). Values with different superscript letters within the same column are significantly different (P < 0.05).
the bitter lupin seed supplement kept MDA lower than that in the hypercholesterolemic diet. Moreover, the bitter lupin seed supplement kept GSH higher than that in the hypercholesterolemic diet. On the other hand, bitter lupin seed supplements improve antioxidant enzymes better than sweet lupin seeds as compared to the hypercholesterolemic control. In general, the lipids profile, transaminases, LDH, MDA of bitter and sweet lupin groups were still higher than in the normal control group, while GSH and enzymic antioxidants were increased for the bitter lupin seed group, they were still lower than those in the normal control group.

Bitter and sweet lupin seeds are natural, normal, healthy and appropriate for reducing oxidative stress, hyperlipidemic, hypertriglyceridemic and hypercholesterolemic factors. These findings stand in stark contrast to the use of hypercholesterolemic drugs that have life-threatening side effects like aching or weakness of skeletal muscles.

Finally the present results clearly illustrate the possibility of using bitter lupin seeds as hypocholesterolemic and antioxidative agents, although further studies using higher concentrations of the seeds may be needed to normalize the rest of the biochemical parameters. It is our opinion, however, that serious experiments must be carried out on human patients.

REFERENCES


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